NEW OXIDIZED ISOCULARINE ALKALOIDS FROM SARCOCAPNOS PLANTS

M.J.Campello(in part), L.Castedo^{*}, D.Domínguez, A.Rodriguez de Lera J.M.Saá, R.Suau, E.Tojo and M.C.Vidal

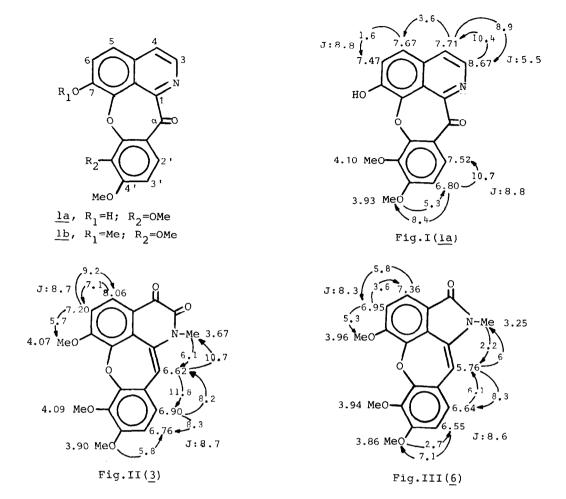
Departamento de Química Orgánica de la Facultad de Química e Instituto de Productos Naturales Orgánicos (Sección Alcaloides) del C.S.I.C.,Santiago (SPAIN)

<u>Abstract</u>: Structures <u>la</u>, <u>3</u> and <u>6</u> were deduced for the new oxoisocularine alkaloids, oxosarcophylline, yagonine and aristoyagonine, respectively, on the basis of spectroscopic studies and synthesis.

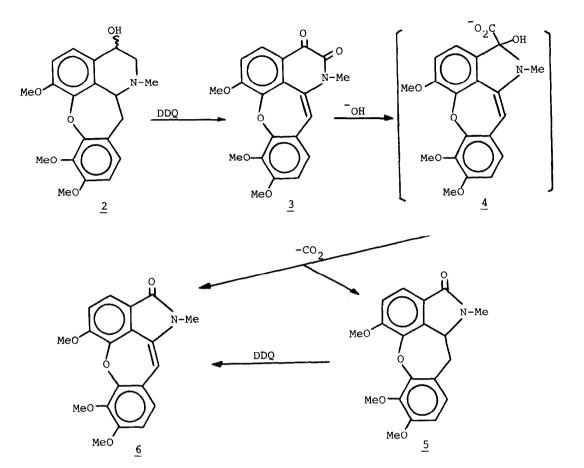
The Fumariaceae plants are a rich source of cularine alkaloids¹. Our current studies on the chemical components of plants of the genus Sarcocapnos, have led us to the isolation of three new cularines, namely oxosarcophylline <u>la</u>, yagonine <u>3</u> and aristoyagonine <u>6</u>, the last two being the first dioxocularine and N-methylated aristocularine, respectively.

Oxosarcophylline la(from S. Enneaphylla (L.)DC and S. Crassifolia (L.)DC) was obtained as yellow needles, mp 170-1719C(EtOH). Its UV spectrum was very similar to that of oxosarcocapnine \underline{lb}^2 , showing absorptions at λ_{max} (EtOH) $(\log \epsilon): 218(4.48), 252(4.27), 330(3.54)$ and 396(3.61) nm, which on addition of acid suffered a bathochromic shift, λ_{max} (EtOH+HCl)(log ϵ):218(4.48), 260(4.24), 395(3.53) and 470(3.25) nm. Its phenolic nature was deduced from a strong bathochromic shift observed on addition of base, λ_{max} (EtOH+NaOH) (log ϵ):218 (4.37), 280(4.25), 310(sh, 3.90) and 510(3.52) nm. Its IR spectrum (KBr) displayed bands at 3400(OH) and 1670 cm⁻¹ (conjugated carbonyl). The molecular formula $C_{1,8}H_{1,3}NO_5$ was established by high resolution MS, which showed the molecular ion at m/e(%):323.0798(100)(calculated:323.0794) and fragments were also observed at m/e 306(34), 295(27), 292(31) and 280(17). The isocularine skeleton was deduced from its PMR spectrum(250MHz,CDCl₃, δ),which exhibited two methoxyl singlets and three aromatic AB quartets. The substitution pattern of oxosarcophylline la was firmely established by O-methylation with diazomethane, which gave a product identical to authentic oxosarcocapnine 1b. The phenolic group was located at the C7 position by comparison of the mass spectrum of oxosarcophylline (which exhibits a low peak at M^+ -43) with those of the other oxocularines which possess a methoxy group at $C_7^{2,3}$. This was confirmed by NOEDS⁴ experiments as shown in Fig.1⁹.

Yagonine 3(from S.Enneaphylla) was obtained as red needles,mp 226-2279C (EtOH). Its UV spectrum showed a highly conjugated system, with absorptions at λ_{max} (EtOH):217,254,340 and 435 nm. Its IR spectrum (KBr) displayed a broad absorption band at 1680 cm⁻¹(C=O), and no signals were apparent at frequencies higher than 3000 cm⁻¹. Its ¹³CNMR spectrum (62.83 MHz,CDCl₃, δ) showed the presence of two carbonyl groups, with signals at 175.24 and 156.96, nine



quaternary sp² carbons (156.76, 155.33, 149.30, 141.98, 141.33, 133.48, 130.03, 122.29 and 121.82), five sp² carbons bound to hydrogen (126.83, 124.26, 118.59, 113.94 and 108.95), three methoxyl groups (61.61, 56.47 and 56.22) and one N-methyl group (32.92). The PMR spectrum, with NOEDS(Fig.II), was consistent with the 3,4-dioxocularine structure <u>3</u>. Its molecular formula $C_{20}H_{17}NO_6$ was confirmed by a high resolution MS which exhibited the molecular ion at m/e(%):367.1061(20)(calculated 367.1056)and fragments were also observed at m/e 366(100), 338(55), 323(57), 308(13) and 280(25). Structure <u>3</u> for yagonine was finally confirmed by comparison with synthetic material, which has been obtained by a method previously reported for the preparation of dioxoaporphines⁵. Thus DDQ oxidation (deoxygenated benzene/809C/2 hrs) of a mixture of the two 4-hydroxysarcocapnine epimers <u>2</u>⁶ afforded yagonine <u>3</u> in 41% yield after column chromatography.



Aristoyagonine 6, (from S. tnneaphylla), was obtained as yellow needles, mp 165-166 \pm C(MeOH). Its UV spectrum showed absorptions at λ_{max} (EtOH):220, 230, 250, 296, 330(sh) and 410 nm (no change upon addition of acid or base was observed). The IR spectrum (KBr) displayed bands at 1700 and 1680 $\rm cm^{-1}$. Its molecular formula $C_{19}H_{17}NO_5$ was confirmed by a high resolution MS, which exhibited the molecular ion at m/e(%):339.1107(M⁺,100)(calculated:339.1107) and fragments were also observed at m/e 324(30), 309(7), 296(10), 281(17), 253(12) and 238(27). Its PMR data (250 MHz,CDCl₃, 6) and NOEDS experiments (Fig.III) suggested the aristoisocularine-type structure $\underline{6}$. The ¹³CNMR spectrum (62.83 MHz, CDCl₃, δ) confirmed the presence of a carbonyl group (166.13), fourteen sp² carbons (154.94, 152.14, 148.05, 141.87, 141.48, 135.42, 127.79, 125.82, 122.03, 121.62, 118.80, 115.03, 108.21 and 107.94), three methoxyl groups (61.19, 56.70 and 56.07) and one N-Me group (25.49). In order to confirm the structure of aristoyagonine $\underline{6}$, we have carried out its synthesis from yagonine 3, by means of a benzilic acid type rearrangment, which has previously been observed to occur easily in the dioxoaporphine alkaloids⁵. However, treatment of a methanolic suspension of yagonine 3 with a large excess of barium hydroxide for four hours at room temperature, gave only a very low yield (7%) of aristoyagonine 6, together with colorless

compound 5^7 (56%). 5 can be assumed to arise from the decarboxylation of the benzilic acid rearranged intermediate 4, which has been isolated in the alkaline treatment of analogous systems⁸. Contrary to what is known in the dioxoaporphines, the oxydized derivative 6 is in our case a minor product in this process. Another noteworthy difference is that no replacement of any methoxyl group was observed when the rearrangment of 3 was carried out with ethanolic sodium hydroxide at room temperature.

Compound 5 was oxidized with DDQ (refluxing benzene, 24 hrs), giving aristoyagonine $\underline{6}$ in quantitative yield, which was identical to the natural compound.

The dioxocularines and aristocularines are probably biogenetically derived from 4-hydroxycularines by further oxidation.

Acknowledgement: We thank the Comisión Asesora (Spain) for its financial support.

REFERENCES AND NOTES

- 1.- For previous work see:
 - a) J.M.Boente, L.Castedo, D.Domínguez, A.Fariña, A.R. de Lera and M.C.
 Villaverde, Tet. Lett., 889 (1984).
 - b) J.M.Boente, L.Castedo, A.R. de Lera, J.M.Saá, R.Suau and M.C.Vidal, 'Tet. Lett., 1829 (1984).
- 2.- M.J.Campello, L.Castedo, J.M.Saá, R.Suau and M.C.Vidal, Tet.Lett., 239(1982).
- 3.- J.M.Boente, L.Castedo, A.R. de Lera, J.M.Saá, R.Suau and M.C.Vidal, <u>Tet. Lett.</u>, <u>24</u>, 2295 (1983).
- 4.- L.D.Hall and J.K.M.Sanders, J. Am. Chem. Soc., 102, 5703 (1980).
- 5.- L.Castedo, R.Suau and A.Mouriño, Tet. Lett., 501 (1976).
- 6.- L.Castedo, D.Domínguez, A.R. de Lera and E.Tojo, <u>Tet. Lett.</u>, submitted for publication.
- 7.- <u>5</u> Crystallized from MeOH (mp 162-164°C) and analysed for $C_{19}H_{19}NO_5$; UV λ_{max} (EtOH):222, 265 and 296(sh) nm; IR(KBr):1680 cm⁻¹; MS: m/e(%): 341(M⁺,100), 326(28), 310(44), 189(28), 176(26) and 167(36); PMR(250 MHz, CDCl₃, δ):3.17(s,3H,-NMe). 3.89(s,3H,-OMe), 4.00(s,3H,-OMe), 4.03(s,3H, -OMe), 4.52, 3.39 and 2.86(ABX,H₁,H_a and H_a respectively, J_{1-aa}=2.7, J_{1-aβ}=11.3, J_{aa-aβ}=13.6), 6.67 and 6.89(ABq, J=8.6,H₃, and H₂,) and 7.05 and 7.52(ABq, J=8.1,H₆ and H₅).
- 8.- P.A.S.Smith and R.O.Kan, J. Am. Chem. Soc., 83, 2580 (1961).
- 9.- Arrows in Figs. I-III have the following meaning: irradiated proton——— % enhancement——— observed proton.

(Received in UK 26 July 1984)